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Mutations in the Pre-Replication Complex cause Meier-Gorlin syndrome

Louise S. Bicknell^{1,19}, Ernie M.H.F. Bongers^{2,19}, Andrea Leitch¹, Stephen Brown¹, Jeroen Schoots², Margaret E. Harley¹, Salim Aftimos³, Jumana Y. Al-Aama^{4,5}, Michael Bober⁶, Paul A. J. Brown⁷, Hans van Bokhoven⁸, John Dean⁹, Alaa Y. Edrees⁵, Murray Feingold¹⁰, Alan Fryer¹¹, Lies H. Hoefsloot², Nikolaus Kau¹², Nine V.A.M. Knoers⁸, James MacKenzie⁷, John M. Opitz¹³, Pierre Sarda¹⁴, Alison Ross⁹, I. Karen Temple¹⁵, Annick Toutain¹⁶, Carol A. Wise¹⁷, Michael Wright¹⁸, and Andrew P. Jackson¹

¹ Medical Research Council (MRC) Human Genetics Unit, Institute of Genetics and Molecular Medicine, Western General Hospital, Crewe Road, Edinburgh, EH4 2XU, UK. ² Department of Human Genetics, Institute for Genetic and Metabolic Disease, Radboud University Nijmegen Medical Centre, PO Box 9101, 6500 HB Nijmegen, The Netherlands. ³ Northern Regional Genetics Service, Auckland Hospital, Auckland, New Zealand. ⁴ Department of Genetic Medicine, Faculty of Medicine, King Abdulaziz University, Jeddah, Saudi Arabia. ⁵ Princess Al Jawhara Center of Excellence in the Research of Hereditary Disorders, King Abdulaziz University, Jeddah, Saudi Arabia. ⁶ Division of Genetics, Department of Pediatrics, A.I. DuPont Hospital for Children, Wilmington, Delaware, USA. ⁷ Department of Pathology, Aberdeen Royal Infirmary, Aberdeen, AB25 9ZD, UK. ⁸ Department of Human Genetics, Nijmegen Centre for Molecular Life Sciences and Institute for Genetic and Metabolic Disease, Radboud University Nijmegen Medical Centre, PO Box 9101, 6500 HB Nijmegen, The Netherlands. ⁹ Department of Clinical Genetics, Ashgrove House, Foresterhill, Aberdeen AB25 2ZA, UK. ¹⁰ National Birth Defects Center, 40, 2nd Avenue, Waltham, MA 02451-1137, USA. ¹¹ Department of Clinical Genetics, Royal Liverpool Children's Hospital, Alder Hey, Eaton Road, Liverpool L12 2AP, UK. ¹² Aberdeen Maternity Hospital, Cornhill Road, Foresterhill, Aberdeen, AB25 2ZL, UK. ¹³ Human Genetics and Obstetrics and Gynecology, University of Utah, Medical Genetics, 50 N. Mario Capecchi Drive, Salt Lake City, UT 84132, USA. ¹⁴ Service de Génétique Médicale, centre de Référence Anomalies du Développement CHRU de Montpellier, Hôpital Arnaud de Villeneuve, 371 avenue du Doyen Gaston Giraud, 34292 Montpellier cedex 1, France. ¹⁵ Division of Human Genetics, University of Southampton, Faculty of Medicine, Southampton General Hospital, Southampton, SO16 6YD,

Correspondence to A.P.J (Andrew.Jackson@hgu.mrc.ac.uk) and E.M.H.F.B (e.bongers@antrg.umcn.nl)..

¹⁹These authors contributed equally

Author Contributions

LSB performed microsatellite genotyping. LSB, SB and JS performed mutation screening of patients and controls with the help of AL, EMHFB, LHH, HvB and NVAMK. MEH performed chromosome breakage analysis. EMHFB and APJ clinically characterized the Meier-Gorlin syndrome cases and performed review of phenotypes and sample collection. SA, JYA-A, MB, PAJB, HvB, JD, AYE, MF, AF, NK, NVAMK, JM, JMO, PS, AR, IKT, AK, CAW and MW contributed clinical cases and clinical data for the study. APJ and LSB wrote the paper with EMHFB.

Competing financial interests

The authors declare no competing financial interests.

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URLs

UniSTS - <http://www.ncbi.nlm.nih.gov/unists>

Exonprimer - <http://ihg2.helmholtz-muenchen.de/ihg/ExonPrimer.html>

ClustalW - <http://www.ebi.ac.uk/Tools/clustalw2/index.html>

CCD - <http://www.ncbi.nlm.nih.gov/cdd>

UK. ¹⁶ Service de Génétique, Hôpital Bretonneau, 2 boulevard Tonnellé, 37044 Tours cedex 1, France ¹⁷ Sarah M. and Charles E. Seay Center for Musculoskeletal Research, Texas Scottish Rite Hospital for Children, Dallas, TX 75219, USA. ¹⁸ Northern Genetics Service, Newcastle Upon Tyne Hospitals NHS Trust, Central Parkway, Newcastle Upon Tyne, NE1 3BZ, UK.

Abstract

Meier-Gorlin syndrome (ear, patella, short stature syndrome) is an autosomal recessive primordial dwarfism syndrome characterised by absent/hypoplastic patellae and markedly small ears¹⁻³. Both pre and post-natal growth are impaired in this disorder and although microcephaly is often evident, intellect is usually normal. We report here that this disorder shows marked locus heterogeneity and we identify mutations in five separate genes: *ORC1*, *ORC4*, *ORC6*, *CDT1* and *CDC6*. All encode components of the pre-replication complex, implicating defects in replication licensing as the cause of a genetic syndrome with distinct developmental abnormalities.

In the accompanying paper⁴ we identified mutations in *ORC1* encoding a subunit of the origin recognition complex (ORC) in five patients with microcephalic primordial dwarfism. The ORC multiprotein complex is loaded onto chromatin at DNA origins prior to S phase to license replication. In conjunction with additional components CDT1, CDC6 and MCM helicase proteins, it then forms the pre-replication complex (described in Fig. 1). Cellular studies established that *ORC1* patients had partial loss-of-function defects in replication licensing⁴. An *orc1* zebrafish model recapitulated the dwarfism phenotype as did morpholino depletion of another pre-replication complex component, *mcm5*. This led us to hypothesise that further pre-replication complex genes might cause primordial dwarfism, however mutations in other ORC subunits were not found.

In this study, the profound growth retardation and microcephaly in a patient with a complex lethal developmental syndrome led us to screen for *ORC1* mutations. We found that he and his brother (P1, P2, Table 1 and Supplementary Table 1) were compound heterozygote for mutations in *ORC1* (p.R105Q and p.V667fsX24). The combination of truncating and missense mutations was in contrast to biallelic missense mutations seen in previous *ORC1* cases, suggestive of greater loss-of-function and consistent with a more severe phenotype. Unlike previous *ORC1* cases, the proband (P1) had an extensive number of developmental malformations including severe cortical dysplasia, congenital emphysema, absence of the pancreatic tail and retroflexion of the knees; as well as micropenis, blepharophimosis and cranial suture stenosis (Fig. 1). Notably, he also had significant microtia and absent patellae, features classically associated with Meier-Gorlin syndrome (MIM 224690).

Consequently, we sequenced *ORC1* in 33 patients with an established diagnosis of Meier-Gorlin syndrome and identified two further cases with mutations in *ORC1* (Table 1, Fig. 2). Both patients from Families 2 and 3 were compound heterozygotes for the same *ORC1* mutations: a splice acceptor site mutation in conjunction with the previously identified c.314G>A that encodes the R105Q amino acid substitution⁴, again in contrast to the biallelic missense mutations observed in the previous study. In Family 2, the c.314G>A mutation was inherited on a haplotype shared with Family 1 (Supplementary Fig. 1), consistent with common ancestry. However, the c.314G>A mutation in Family 3 and the two splice acceptor site mutations in Family 2 and 3 appear to have arisen independently. Notably, P3 in Family 2 is the original patient described by Gorlin², unequivocally establishing that mutations in *ORC1* cause Meier-Gorlin syndrome.

Sequencing the genes encoding other ORC subunits then identified pathogenic mutations in two further subunits of the Origin Recognition Complex (*ORC4*, *ORC6*) in three patients

with Meier-Gorlin syndrome (Table 1, Supplementary Table 1). Monozygotic twins P6 and P7 were found to be homozygous for a missense mutation (c.521A>G, p.Y174C) in *ORC4* (family also independently ascertained by M. Samuels and colleagues, ref. ⁵). The substituted tyrosine was conserved throughout evolution as distant as *S. cerevisiae* and *S. pombe*, and was located within Box VI of the AAA domain⁶ (Fig. 2), a domain that is essential for ATP-dependent complex formation of ORC4 with other ORC subunits^{7,8}. A second patient, P5, is a compound heterozygote for this same Y174C substitution (present on the ancestral haplotype shared with P6 and P7; Supplementary Fig. 1), in conjunction with a four base-pair insertion mutation causing a translational frameshift and premature protein termination.

In Family 6, three affected children were compound heterozygotes for mutations in *ORC6*. Similar to P5, these patients inherited one loss-of-function mutation caused by a two base pair deletion and a missense mutation resulting in the substitution (p.Y232S) of an amino acid completely conserved from fungi to humans located at the C-terminus of the ORC6 protein (Fig. 2).

Given that mutations in genes encoding multiple components of the ORC complex cause Meier-Gorlin syndrome, we next examined whether additional components of the pre-replication complex could be similarly implicated. CDT1 and CDC6 are recruited to the pre-replication complex following assembly of the full ORC complex on chromatin⁹. As the region mutated in ORC6 is involved in CDT1 protein binding in *S. cerevisiae*¹⁰, we next screened our Meier-Gorlin syndrome cohort for mutations in *CDT1*. Five families were identified with compound heterozygous mutations in *CDT1* (Table 1). In three of these families, one of the mutations was c.1385G>A, causing the substitution R462Q in the C-terminal winged helix domain of CDT1 (Fig. 2). This residue has been implicated in MCM helicase complex binding through mutagenesis of the orthologous mouse residue (p.R474Q)¹¹, the same substitution as the human mutation we have identified here.

Finally, screening of *CDC6* identified one Meier-Gorlin syndrome patient (P18) homozygous for a missense mutation (c.968C>G), resulting in the substitution T323R. This residue is absolutely conserved from fungi to humans (Fig. 2) and lies within the box VII motif of the AAA domain (Fig. 2)⁶, an ATP binding domain that is essential for CDC6 function in DNA replication¹².

We report here that mutations in multiple components of the pre-replication complex cause Meier-Gorlin syndrome and despite finding marked genetic heterogeneity, identify an underlying functional homogeneity in this disorder. The functional studies of ORC1 in the accompanying manuscript⁴ establish that impairment of ORC1 function is associated with extreme growth failure in humans. Here, our molecular genetic findings implicate the entire pre-replication complex in primordial dwarfism syndromes, providing compelling genetic evidence supporting the notion that impaired replication licensing causes growth failure.

Though most of the patients described in this paper have typical features of Meier-Gorlin syndrome¹⁻³, considerable phenotypic variation is seen from mutations in these genes, particularly with the ORC1 cases reported here and in the accompanying paper⁴. Growth parameters vary markedly (height ranges from -0.4 to -9.6 SD, and head circumference from +1.7 to -9.8 SD), with cognition varying from normal to severe neurological impairment associated with a severe cortical dysplasia. Thus, although microtia and small patellae are the best predictors of mutations in the pre-replication complex, the disease spectrum associated with these genes could extend from cases described as microcephalic osteodysplastic primordial dwarfism I/III to those with nonsyndromic primordial dwarfism. (Further discussion of clinical phenotypes, see Supplementary Note.)

Impaired replication might be expected to affect growth of all tissues equally; therefore it is noteworthy that specific tissues are disproportionately reduced in size, most readily apparent in the ears and patellae. Most strikingly, significant congenital malformations are also evident in several patients, affecting limb, genitourinary, lung and brain development. This finding surprisingly links mutations in a chromatin bound protein complex regulating replication with multiple congenital malformations. Such an association between chromatin bound complexes involved in essential cellular functions and development is not unprecedented. Notably in Cornelia de Lange syndrome (MIM 122470), mutations in the cohesion complex also cause severe growth retardation though with very different developmental consequences¹³.

The cellular mechanisms, by which preRC components cause the specific developmental phenotypes reported here, are not immediately evident. It may be that proliferation of certain cell types, such as chondrocytes in the ears and patellae are particularly sensitive to reduced preRC function. Alternatively, these and the other developmental anomalies could result from other cellular functions of the preRC components, as has been found for individual ORC proteins¹⁴.

Previous primordial dwarfism genes have DNA damage response and centrosome functions¹⁵⁻¹⁷, while here we find mutations in genes involved in licensing of DNA replication origins. All are critical components required for cell cycle progression. Pathogenesis is therefore likely to be the result of impaired cellular proliferation, resulting in reduced cell number and consequently global growth failure. While in this framework genes regulating replication are rational candidates for primordial dwarfism, it is most surprising to find specific congenital malformations occurring as the consequence of mutations in the pre-replication complex. This suggests an unanticipated link between the regulation of replication and development that warrants future investigation.

Methods

Research subjects

All patients in this study had previously been published as cases of Meier-Gorlin syndrome or fulfilled diagnostic criteria for Meier-Gorlin syndrome (microtia, absent/small patellae and short stature). Genomic DNA was isolated from peripheral blood of the affected children and family members. No cytogenetic abnormalities were observed on routine karyotyping of these Meier-Gorlin syndrome cases (n=10 patients). Furthermore, no significant increase of either spontaneous, or radiation-induced chromosome breaks was observed (2 cases versus 2 control patients, 100 metaphases assessed per patient; $p = 0.7$, X^2 -test). Informed consent was obtained from all participating families and the studies were approved by the Scottish Multicentre Research Ethics Committee (04:MRE00/19), Regional Committee on Research Involving Human Subjects Nijmegen - Arnhem (0006-0119); or University of Texas Southwestern Medical Center at Dallas (IRB #032008-066). Informed consent was obtained for the publication of photographs.

Sequencing of candidate genes

Primers were designed using Exonprimer (Supplementary Table 2). Coding exons and exon-intron boundaries of each gene were screened by bidirectional capillary sequencing on an ABI 3730 gene sequencer. Primer sequences and PCR conditions are available upon request. Sequence analysis and mutation detection performed using Mutation Surveyor v. 2.61 (Softgenetics LLC).

Microsatellite Genotyping

Microsatellites were selected on the basis of heterozygosity and proximity to each gene. Primer sequences for microsatellites were obtained from UniSTS. Amplicons were separated by capillary electrophoresis on an ABI3730 Sequencer, and allele sizes obtained using GeneMapper (Applied Biosystems).

Protein alignments

Reference sequences for all proteins were obtained from RefSeq and alignments performed using ClustalW2. Protein domains were mapped using data obtained from Conserved Domain Database (CCD).

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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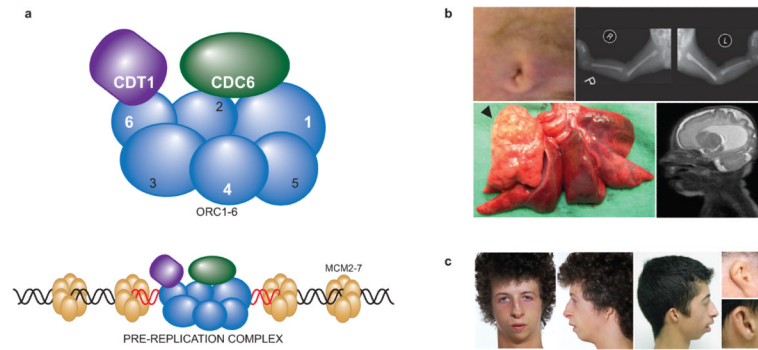


Figure 1. The pre-replication complex and Meier-Gorlin syndrome

(a) Genome replication is licensed by the binding of a number of specialised proteins to origins of replication, which form the pre-replication complex⁹. The first step in complex formation is the loading of the heterohexameric Origin Recognition Complex (comprising ORC1-6 proteins), onto chromatin in an ATP dependent manner during M and G1 phases of the cell cycle. Further proteins, including CDC6 and CDT1 are then recruited to the pre-replicative complex, that then permit reiterative loading of the multimeric MCM helicase. At the commencement of S-phase, replication is started by the MCM helicase unwinding DNA, and the recruitment of additional replication proteins. The five proteins implicated in Meier-Gorlin syndrome are highlighted in white text (ORC1, ORC4, ORC6, CDT1, CDC6). (b) Patient 1 has a severe developmental malformation syndrome with marked microtia and extreme retroflexion and dislocation of the knees (top row). His malformations included lobar congenital emphysema (arrow head), and a severe cortical dysplasia of the brain. Parasagittal T2 weighted MRI at age 1 month demonstrating severe pachygyria, most severe frontally along with ventricular enlargement. (c) Two patients (P11, P9) with classical Meier-Gorlin syndrome. **b,c** Informed consent to publish the photographs was obtained from the subjects' parents.

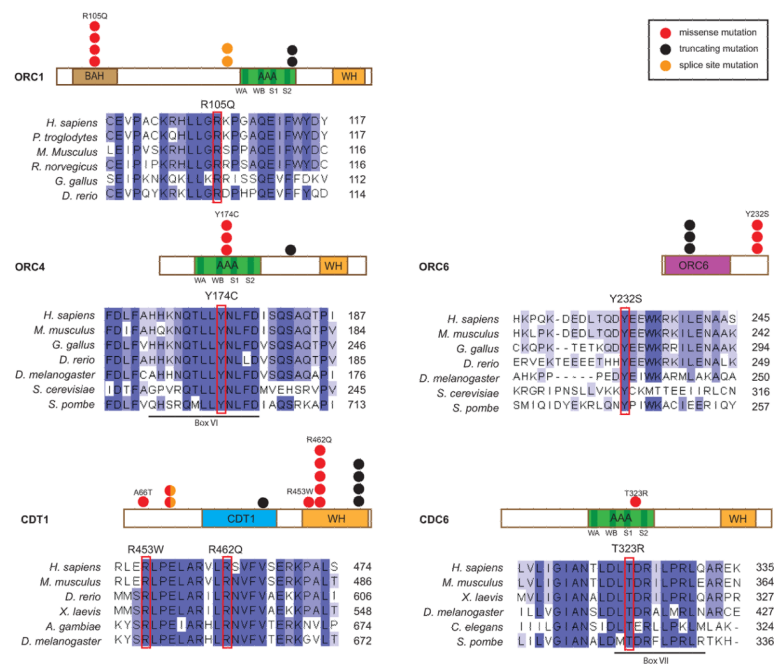


Figure 2. Pre-replication complex proteins mutated in Meier-Gorlin syndrome

Schematics for each protein depicting known protein domains with positions of mutations shown by filled circles. Each filled circle represents one patient. Clustalw alignment of protein residues surrounding substituted amino acids (positions of substituted residues indicated by red boxes). WA, Walker A; WB Walker B, S1, Sensor 1; S2, Sensor 2; motifs. BAH, Bromo Associated Homology domain; AAA, ATPase Associated with a wide range of cellular Activities; WH, winged helix domain.

Table 1
Mutations in five genes encoding pre-replication complex proteins in patients with Meier-Gorlin syndrome.

Family	Patient	Ancestry	Gene	Nucleotide Alterations	Amino Acid Alterations	Exon(s)	Segregation	Parental Consanguinity	Gender	Current Height (SD)	Current OFC (SD)	Microtia	Absent/Small Patellae
F1	P1	UK	<i>ORC1</i>	[c.314G>A] + [c.1999_2000delGTinsA]	R105Q + V667fsX24	4; 13	Het, M, P	No	M	-9.6	-9.8	+	+
F1	P2	UK	<i>ORC1</i>	[c.314G>A] + [c.1999_2000delGTinsA]	R105Q + V667fsX24	4; 13	Het, M, P	No	M	NA	NA	+	u
F2	P3	USA	<i>ORC1</i>	[c.314G>A] + [c.1482-2A>G]	R105Q + intron 9 splice acceptor site	4; intron 9	Het, M, nps	No	M	-6.6	u	+	+
F3	P4	UK	<i>ORC1</i>	[c.314G>A] + [c.1482-2A>G]	R105Q + intron 9 splice acceptor site	4; intron 9	Het, M, P	No	F	-6.9	-4.0	+	+
F4	P5	USA	<i>ORC4</i>	[c.521A>G] + [c.874_875insAACAA]	Y174C + A292fsX19	8; 11	Het, M, P	No	F	-6.4	u	+	+
F5	P6	USA	<i>ORC4</i>	c.521A>G	Y174C	8	Hom, M, P	Yes	F	-4.2	-2.1	+	-
F5	P7	USA	<i>ORC4</i>	c.521A>G	Y174C	8	Hom, M, P	Yes	F	-4.1	-3.0	+	-
F6	P8	TR	<i>ORC6</i>	[c.257_258delTT] + [c.695A>C]	F86X + Y232S	3; 7	Het, M, P	Yes	F	-3.3	-1.6	+	+
F6	P9	TR	<i>ORC6</i>	[c.257_258delTT] + [c.695A>C]	F86X + Y232S	3; 7	Het, M, P	Yes	M	-2.4	-2.1	+	+
F6	P10	TR	<i>ORC6</i>	[c.257_258delTT] + [c.695A>C]	F86X + Y232S	3; 7	Het, M, P	Yes	M	-3.2	-2.3	+	+
F7	P11	NZ	<i>CDT1</i>	[c.1385G>A] + [c.1560C>A]	R462Q + Y520X	9; 10	Het, M, P	No	M	-4.7	+0.1	+	+
F8	P12	UK	<i>CDT1</i>	[c.196G>A] + [c.351G>C]	A66T + Q117H(exon 2 splice donor site)	1; 2	Het, M, P	No	F	-5.1	-5.0	+	+
F9	P13	USA	<i>CDT1</i>	[c.1385G>A] + [c.1560C>A]	R462Q + Y520X	9; 10	Het, M, P	Yes	F	-4.7	-1.3	+	+
F9	P14	USA	<i>CDT1</i>	[c.1385G>A] + [c.1560C>A]	R462Q + Y520X	9; 10	Het, M, P	Yes	F	-3.9	-1.0	+	+
F9	P15	USA	<i>CDT1</i>	[c.1385G>A] + [c.1560C>A]	R462Q + Y520X	9; 10	Het, M, P	Yes	M	-1.6	+1.7	+	+
F10	P16	UK	<i>CDT1</i>	[c.351G>C] + [c.1385G>A]	Q117H(exon 2 splice donor site) + R462Q	2; 9	Het, nps	Yes	F	-3.3	-0.5	+	+
F11	P17	SA	<i>CDT1</i>	[c.1081C>T] + [c.1357C>T]	Q361X + R453W	7; 9	Het, M, P	No	F	-0.4	-2.1	+	+
F12	P18	FR	<i>CDC6</i>	c.968C>G	T323R	7	Hom, M, P	Yes	M	-4.1	-3.3	+	+

Mutations are described numbered from the first nucleotide of the initiation codon in the nucleotide sequence. For each mutation, 380 control chromosomes were screened and found to be negative for the sequence change. Abbreviations: Hom, homozygous in affected individual; Het, compound heterozygous in affected individual. M, mutation identified in mother; P, mutation identified in father; nps, parental sample(s) not available. The protein truncation Y520X in *CDT1* is within the last exon and therefore the transcript is not expected to undergo nonsense-mediated mRNA decay. Additionally, in *CDT1* the mutation c.351G>C confers a non-conservative amino acid substitution (p.Q117H), however the nucleotide lies within the exon 2 splice donor site and therefore this mutation is expected to result in an inframe deletion of exon 3 from the transcript and resulting protein. TR, Turkey; NZ, New Zealand; FR, France; SA, Saudi Arabia; SD, standard deviation, OFC, occipito-frontal circumference; NA, not available; u, unknown.